

REVIEW

Use and adequacy of non-pregnancy diagnosis in cow. Which future?

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Abstract

In cattle, early detection of gestation is very important from an economic and management point of view in all types of farming. However, due to the poor efficiency of oestrous detection, it is essential to determine non-pregnant cows as early as possible, in order to minimize the inter-insemination interval, thus de facto, reducing herd open days. Direct and indirect gestation diagnostic methods have been developed with the aim of improving the reproductive performance of the herd. Today, the most accurate method for making an early diagnosis of gestation from 28 to 30 days post-insemination is B-mode ultrasound. In recent years, indirect methods have included techniques that allow non-pregnant cows to be identified with a minimum margin of error, the most widely utilized of which is the colour Doppler. This technique is rapidly becoming established for the diagnosis of non-pregnancy, which allows for the identification of non-pregnant animals earlier compared with the pregnancy diagnosis. Some limitations of this technique in dairy cow have been presented.

KEYWORDS

bovine, Doppler, pregnancy, reproductive management, ultrasonography

1 | INTRODUCTION

In the mid-1980s, B-mode ultrasonography began to be used as a complementary examination in bovine reproductive management (DesCôteaux et al., 2009). This technique was initially used for pregnancy diagnosis (Photo 1) (Fissore et al., 1986; Szenci et al., 1999) but soon was utilized to study follicular dynamics, corpus luteum (Acosta & Miyamoto, 2004; Kito et al., 1986), embryonic and foetal development (Curran et al., 1986; Kraison et al., 2018), uterine pathologies (Fissore et al., 1986; Meira et al., 2012), gestational losses (Chaffaux et al., 1986; Chaudhary & Purohit, 2012) foetal sexing (Photo 2) (Ali & Fahmy, 2008; Fricke, 2002) physio-pathology of ovary (Photo 3) (Díaz et al., 2019; Gnemmi, 2001; Quintela et al., 2012) and uterus (Bollwein et al., 2002; Debortolis et al., 2016; Sharma et al., 2019) (Photo 4), diagnosis of twin gestation (Photo 5) (Colloton et al., 2010; López-Gatius et al., 2017; Silva del Rio et al., 2009), diagnosis of embryo and foetal death (Photo 6) (Colloton et al., 2010; Gnemmi, 2004b), diagnosis of foetal malformation (Photo 7)

(DesCôteaux et al., 2010; Gnemmi, 2004b; Gorjidoz et al., 2021) diagnosis of gestational diseases (Photo 8) (DesCôteaux et al., 2009; Murakami et al., 2019) and recently also to improve the efficiency of selecting donor cows/heifers for embryos and oocytes and to improve efficiency of selecting recipients for embryo transfer (Pugliesi et al., 2018; Stroud & Durocher, 2010).

More recently ultrasonography, thanks to the use of the colour Doppler, is becoming an important tool for diagnosis of non-pregnancy diagnosis (NPD) (Photo 9) (Siqueira et al., 2013), which means to predict the absence of a pregnancy before that the pregnancy diagnosis (PD) itself can be achieved and is based on an indirect assessment of the level of progesterone produced by the corpus luteum by measuring its vascularization surface.

Colour Doppler ultrasonography is accurate to diagnose NPD cows and heifers as early as 20 days after post-insemination (Guimaraes et al., 2015; Siqueira et al., 2013; Viana et al., 2013) or 14 days after embryo transfer (ET) (Palhão et al., 2020; Pugliesi et al., 2014) while pregnancy diagnosis (PD) can be applied at day

28–30 post-insemination (Munoz del Real et al., 2006; Pierson & Ginther, 1984). These differences have an important impact on the income of dairy and beef farming because a 30-day of lost gestation, regardless of the number of lactations and days of lactation, costs about \$300 (Cabrera, 2012; De Vries, 2006) and to lose a gestation during the period of 30 to 60 days costs approximately \$600. (De Vries, 2006). Thus, the diagnosis of NPD has a fundamental economic significance, because allows the re-insemination of non-pregnant cows earlier compared with the PD (Andrade et al., 2019; Sá Filho et al., 2014). In fact, with NPD is possible to re-inseminate beef cows at day 25 after the previous insemination compared the 32–34 days of the traditional PD (Palhão et al., 2020). In dairy cows, animals diagnosed as non-pregnant at 32 days post-insemination can be re-inseminated at 35 days if they have a corpus luteum and at 42 days post-insemination if they do not have a corpus luteum at the time of PD negative (Sauls & Stevenson, 2017; Wijma et al., 2017).

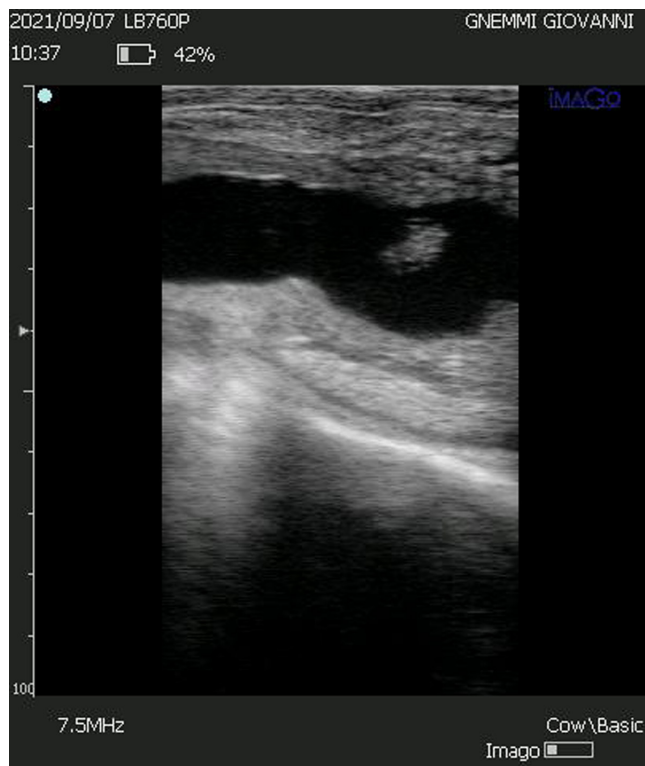


PHOTO 1 Pregnancy diagnosis carried out at 30 days post-insemination in B-mode (echography image by Dr. G. M. Gnemmi)

Therefore, the aim of this article is to describe the NPD by echo Doppler in cows and analyse the advantages and limitations of this technique with particular attention from the point of view of the veterinary gynaecologist working in the field.

1.1 | Determination of pregnancy (PD) and non-pregnancy (NPD) in cow

In cattle, the determination of pregnancy can be done directly or indirectly. Direct methods are based on the identification of the embryo or foetus (Ginther, 1995; Gnemmi, 2004a) and include techniques such as balloting, rectal palpation and B-mode ultrasound diagnosis (Balhara et al., 2013; Christiansen, 2015; Colloton et al., 2010) together with checking of the ovaries for the presence of one or more corpus luteum (CL) and the uterine horns for the presence of fluids. All the above systems can give a good accuracy of pregnancy from day 30 of gestation onwards (Colloton et al., 2010; Fricke et al., 2016; Kastelic et al., 1989). In some reports, has been indicated the possibility to apply transrectal B-mode ultrasound diagnosis at day 25–30 post-insemination with a satisfactory rate of accuracy (Bollwein et al., 2016; Fricke et al., 2002) but not in others (Badtram et al., 1991; Cabrera, 2012; Romano et al., 2006) because there is a quite high rate of false negative (for review see Szenci, 2021). In conclusion, the PD with B-mode ultrasound cannot be applied in field conditions before day 25–30 from insemination. (Gnemmi & Maraboli, 2008).

Other possibility to achieve an earlier pregnancy diagnosis is by the dosage of progesterone in milk and blood (Chebel et al., 2003; Faustini et al., 2007; Martins et al., 2018), by measuring the glycoproteins of gestation (Balhara et al., 2013; Reese et al., 2018; Ricci et al., 2015) or, more recently, by the evaluation of the CL vascularization by the colour Doppler (Siqueira et al., 2013, 2019; Utt et al., 2009). The determination of pregnancy by plasma or milk level of progesterone or glycoprotein plasma levels has a limit that can be still elevated even in non-pregnant cattle, or in animals that have lost gestation from 7 to 14 d after pregnancy loss (Ricci et al., 2015) with the risk of false positive PD. Indeed, when opting for an early diagnosis of pregnancy, it is essential to ensure that the method has a very low false negative rate (Andrade et al., 2019; Pugliesi et al., 2017). The absence of false negative is a key due to the abortion risk when there is the use of hormonal treatments like resynch in cows that are

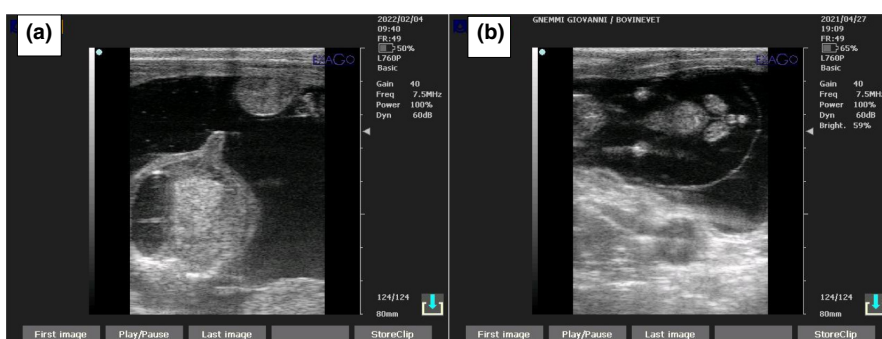


PHOTO 2 Diagnosis of foetal sexing by B-mode ultrasound. (a) Male; (b) female (echography image by Dr. G. M. Gnemmi)

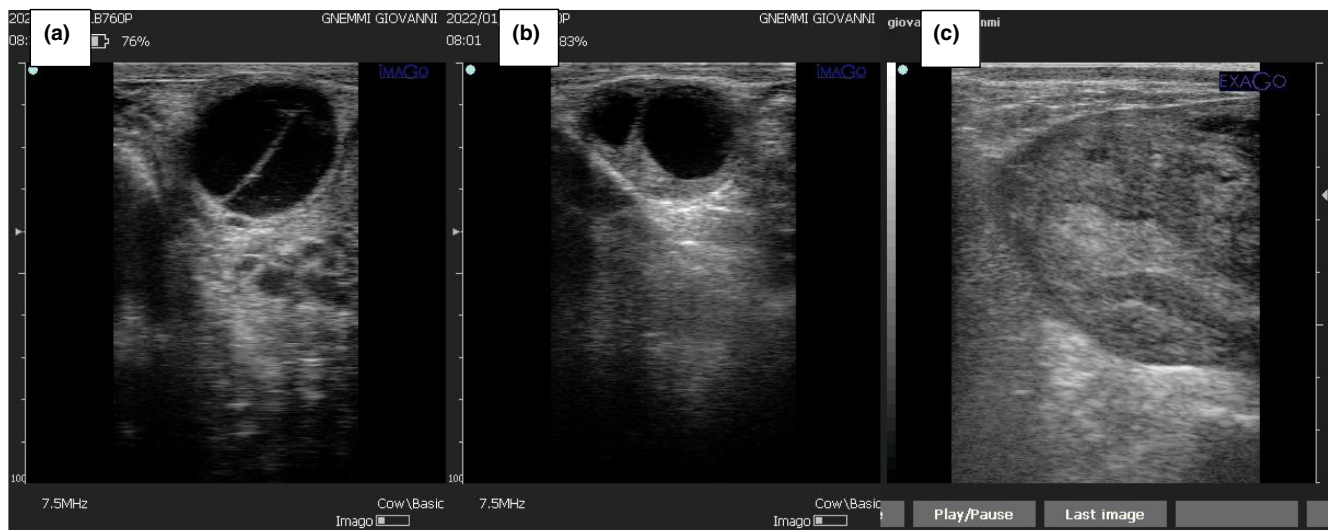


PHOTO 3 B-mode ultrasound diagnosis of ovarian diseases. (a) Follicular cyst; (b) luteal cyst; (c) granulosa cell's tumour (echography image by Dr. G. M. Gnemmi)

pregnant but that are misdiagnosed as being non-pregnant (Pugliesi et al., 2017).

1.1.1 | The colour Doppler application for non-pregnancy diagnosis

Thanks to the colour Doppler, it is possible to evaluate changes of blood flow in CL and therefore the evaluation of functional regression of CL. In this way is possible to predict which animals are not pregnant (NPD diagnosis) by day 18–23 after insemination (Giordano et al., 2012; Siqueira et al., 2013; Utt et al., 2009) or even earlier on day 13 post-insemination (Palhão et al., 2020). In fact, the functional regression of CL anticipates approximately 60–72 hours of the CL anatomical regression (Ginther, 1995; Miyamoto et al., 2005). Thus, at 18–19 days post-insemination, by the evaluation of the functional but not the anatomical regression of CL is possible to predict whether the cow is pregnant or not (Ginther, 2007).

During the meta-estrus, most luteal cells make contact with one or more capillaries, making the CL one of the most vascularized structures in the body (Miyamoto et al., 2005) and at this phase of oestrous cycle, vascular endothelial cells account for up to 50% of the structure of CL (Lei et al., 1991; Miyamoto et al., 2005; O'Shea et al., 1989) and plasma progesterone concentration increases linearly reaching its maximum level on day 14 of the oestral cycle (7.0–8.0 ng/ml) (Herzog et al., 2010).

In a manner mirroring the luteal blood flow increase, plasma progesterone levels increase reaching a maximum level on day 14 and 16 of the oestral cycle (Table 1; Herzog et al., 2010). In the absence of maternal recognition of gestation, cells of the endometrial epithelium produce $\text{PGF2}\alpha$, the main luteo-lysin in the cow (Miyamoto et al., 2005). $\text{PGF2}\alpha$ triggers a series of vascular and hormonal processes leading to luteolysis. During the first 3 days of the follicular phase of the cycle (pro-estrus), the luteal blood flow and plasma

progesterone (<1.0 ng/ml) concentration are drastically reduced (Herzog et al., 2010).

Despite the different opinions on when to make the NPD using colour Doppler, most of the studies suggest that the use of Doppler for the diagnosis of NPD is most accurate when performed between days 18–21 post-insemination. During this time, in non-pregnant cows, there is a reduction in the CL blood flow that induces a drastic drop in progesterone (Bollwein et al., 2016; Lüttgenau & Bollwein, 2014; Scully et al., 2014).

Thus, echo Doppler is a valuable tool for NPD, but there are some limitations that need to be taken into account:

1. At the timing of luteolysis, there are differences between cows with two follicular waves and cows with three follicular waves. 20% of dairy cows and 80% of beef cows have three waves of follicular growth (Adams et al., 2008). Indeed, the number of follicular waves determines the time at which luteolysis takes place and this determines the length of the oestrous cycle (Adams et al., 2008). Therefore, in two-wave cows, complete luteolysis takes place around day 16 of the cycle, while in three-wave cows, luteolysis takes place around day 19 of the cycle. (Herzog et al., 2010). As consequence, the time of the formation and regression of CL is correlated with the pattern of follicular waves (two or three). When a diagnosis of non-gestation is made by colour Doppler between day 18 and 20 post-insemination, there is the risk of finding, in non-pregnant cows, a CL with a vascularization area of 0.3–0.7 cm^2 ($\geq 25\%$ peripheral vascularization of the CL) suggesting a pregnancy (Herzog et al., 2010). Therefore, the accuracy of the diagnostic can be reduced due to false positives (Utt et al., 2009).
2. It should not be underestimated that there is an individual difference regarding the vascularization surface of the CL, which further complicates the definition of the best time to perform this



PHOTO 4 B-mode ultrasound diagnosis of mucopurulent endometritis (echography image by Dr. G. M. Gnemmi)



PHOTO 6 B-mode ultrasound diagnosis of embryo death (30 days post-insemination) (echography image by Dr. G. M. Gnemmi)

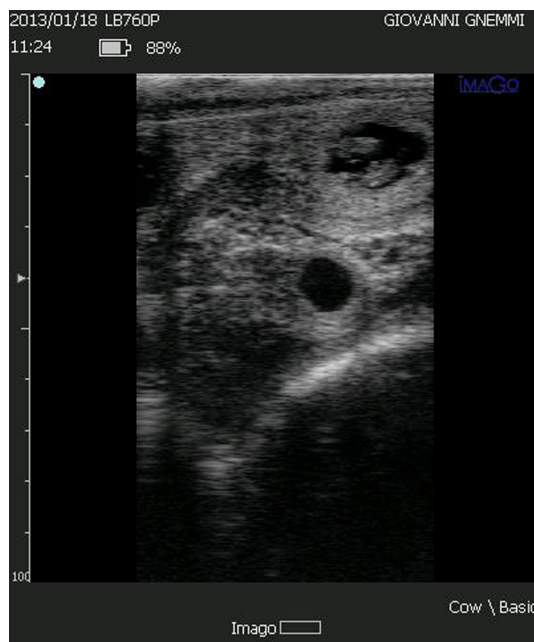


PHOTO 5 B-mode ultrasound diagnosis of twins (echography image by Dr. G. M. Gnemmi)



PHOTO 7 B-mode ultrasound diagnosis of foetal malformation (*Schistosomus reflexus*) (echography image by Dr. G. M. Gnemmi)

examination (Herzog et al., 2011; Lüttgenau & Bollwein, 2014; Utt et al., 2009).

- In high-producing dairy cows, there is a high incidence of the elongated luteal phase (Giordano et al., 2012; Ranasinghe et al., 2011). In these cattle, even in absence of pregnancy, it is normal to find an extensively vascularised CL, even beyond the 20th day post-insemination.
- In high-producing dairy cows, most embryo losses occur between day 6 and 25 of gestation. When embryo losses occur at day 13–14 post-insemination, luteolysis is delayed, and therefore, it is normal to see a widely vascularized CL in these cows, even beyond day 20 post-insemination (Diskin et al., 2006; McNeill et al., 2006).

- In inseminated cows with subclinical endometritis, or in cows developing endometritis after insemination, a persistent CL may be even in the absence of maternal recognition of gestation (Peter et al., 2009). In these cows, the high levels of PGE2 produced by the endometrial stroma cells block the luteolysis (Mwaanga & Janowski, 2000).

Despite these limitations, the colour Doppler for the diagnosis of NPd between 18 and 21 days is interesting, and the accuracy can be improved by the combination with colour Doppler ultrasound assessment of uterine stratigraphy and by the evaluation of the levels

PHOTO 8 B-mode ultrasound diagnosis of pathologies of the gestation. (a) Maceration; (b) mummification (echography image by Dr. G. M. Gnemmi)

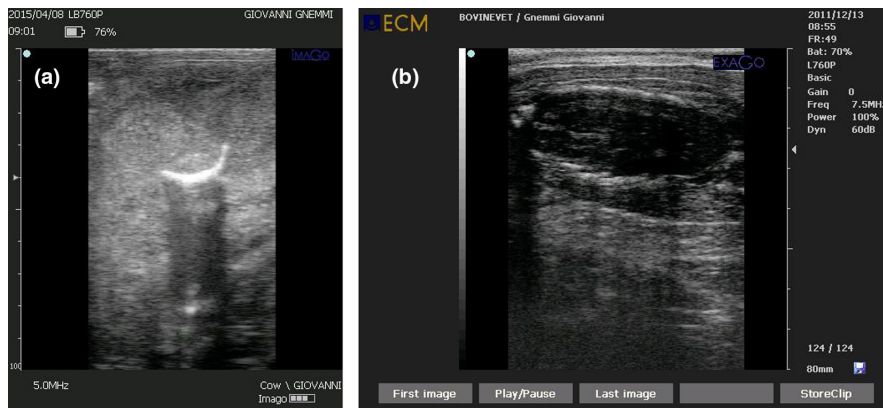


PHOTO 9 Evaluation of LBF. (a) LBF in a cow with progesterone >1.0 ng/ml; (b) LBF in a cow with progesterone <1.0 ng/ml (echography image by Dr. G. M. Gnemmi)

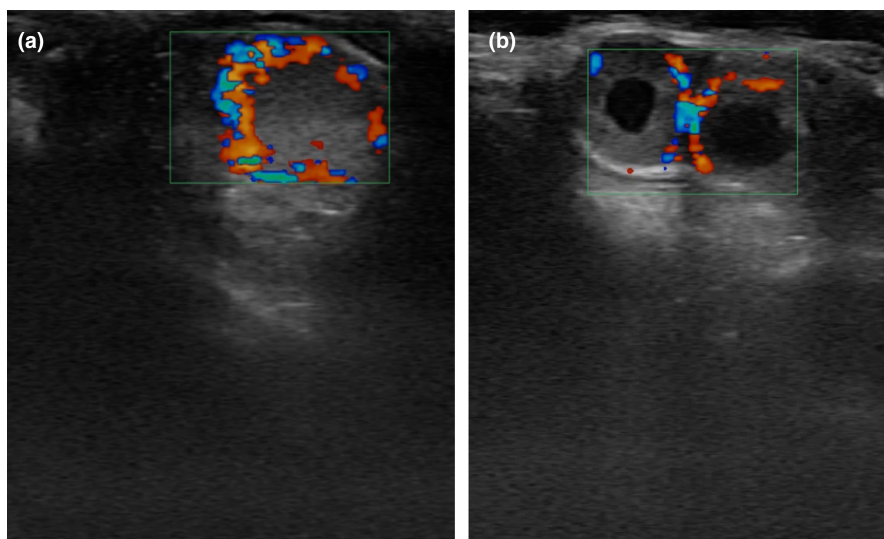


TABLE 1 Progesterone level (ng/ml) and CL vascularization surface area (cm²), depending on the days of the oestral cycle (processed from Herzog et al., 2010)

Day	P4 (ng/ml)	LBS (cm ²)
4	1.7	0.3
7	4.0	0.7
14	7.3	1.3
16	7.1	1.2
17	5.4	1.0
18	2.7	0.7
19	0.8	0.2
20	0.6	0.1
21	0.5	0.1

of hormones (i.e progesterone), to reduce the margin of uncertainty of NPD.

2 | CONCLUSION

The US has given new insight to determine whether a cow becomes pregnant or not. The B-mode Doppler can allow to individuate the

embryo, and this method is the most reliable, but not before day 28–30 of gestation. Conversely, the NPD that is based on the evaluation of CL vascularization by Colour Doppler, allows to identify the cows that are not pregnant before day 28. However, there are some limitations in the use of Colour Doppler for NPD, like the individual difference in the CL vascularization surface, the pattern of follicular waves (two or three), the development of persistent CL or embryo losses.

NPD is important because opened up an excellent opportunity by allowing these cows to be re-inseminated as early as 25 days after the last insemination by the use of resynchronization protocols in this strategy. All inseminated cows receive a resynchronization protocol at around day 22 after AI on day 30. The protocol is cancelled for those diagnosed as pregnant, or completed in non-pregnant ones, and re-insemination occurs within 32 days after the first FTAL (Giordano et al., 2012; Palhão et al., 2020; Sauls & Stevenson, 2017).

Overall, the NPD technique presents some limits and in large commercial dairy herds, it must be carefully taken into account the cost/benefit of this technique before being utilized. Conversely, NPD can be fully justified in small organic dairy farms where is possible for an individual approach to the reproductive management.

The progress of technology may make it possible to create ultrasound units with built-in artificial intelligence to automatically

translate the vascular surface of the corpus luteum into ng/ml of progesterone. So, the NPD would then be simpler and of more practical application for the routine reproduction management of the farm.

AUTHOR CONTRIBUTIONS

Our paper is a review and all the authors have been involved in it with a special role for the section of the review in which the author is expert. Furthermore, Gnemmi G decided the topic, proposed the chapters and organized the manuscript; Fabio De Rensis and Roberta Saleri contributed to the overall review of the manuscript; Gnemmi Benedetta and Maraboli contributed to the basic organizations of the manuscript and literature review.

CONFLICT OF INTEREST

There is no conflict of interest, in accordance with the guidance for Authors provided by the publishing house.

DATA AVAILABILITY

The data that support the findings of this study are available on request from the corresponding author. The data are not publicly available due to privacy or ethical restrictions.

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REFERENCES

- Acosta, T. J., & Miyamoto, A. (2004). Vascular control of ovarian function: Ovulation, corpus luteum formation and regression. *Animal Reproduction Science*, *82*–83, 127–140.
- Adams, G. P., Jaiswal, R., Singh, J., & Malhi, P. (2008). Progress in understanding ovarian follicular dynamics in cattle. *Theriogenology*, *69*, 72–80.
- Ali, A., & Fahmy, S. (2008). Ultrasonographic fetometry and determination of fetal sex in buffaloes (*Bubalus bubalis*). *Animal Reproduction Science*, *106*, 90–99.
- Andrade, J. P. N., Andrade, F. S., Guerson, Y. B., Domingues, R. R., Gomez-León, V. E., Cunha, T. O., Jacob, J. C. F., Sales, J. N., Martins, J. P. N., & Mello, M. R. B. (2019). Early pregnancy diagnosis at 21 days post artificial insemination using corpus luteum vascular perfusion compared to corpus luteum diameter and/or echogenicity in Nelore heifers. *Animal Reproduction Science*, *209*, 106–144.
- Badtram, G. A., Gaines, J. D., Thomas, C. B., & Bosu, W. T. K. (1991). Factors influencing the accuracy of early pregnancy detection in cattle. *Theriogenology*, *35*, 1153–1167.
- Balhara, A. K., Gupta, M., Singh, S., Mohanty, A. K., & Singh, I. (2013). Early pregnancy diagnosis in bovines: Current status and future directions. *The Scientific World Journal*, *2013*, 1–11.
- Bollwein, H., Baumgartner, U., & Stolla, R. (2002). Transrectal Doppler sonography of uterine blood flow in cows during pregnancy. *Theriogenology*, *57*, 2053–2061.
- Bollwein, H., Heppelmann, M., & Lüttgenau, J. (2016). Ultrasonographic Doppler use for female reproduction management. *Veterinary Clinics: Food Animal Practice*, *32*, 149–164.
- Cabrera, V. E. (2012). A simple formulation and solution to the replacement problem: A practical tool to assess the economic cow value, the value of a new pregnancy, and the cost of a pregnancy loss. *Journal of Dairy Science*, *95*, 4683–4698.
- Chaffaux, S., Reddy, G. N. S., Valon, F., & Thibier, M. (1986). Transrectal real-time ultrasound scanning for diagnosing pregnancy and for the monitoring embryonic mortality in dairy cattle. *Animal Reproduction Science*, *10*, 193–200.
- Chaudhary, A. K., & Purohit, G. N. (2012). Ultrasonographic detection of early pregnancy loss in dairy cows. *Journal of Animal Science Advances*, *2*, 706–710.
- Chebel, R. C., Santos, J. E. P., Cerri, R. L. A., Galvao, K. N., Juchem, S. O., & Thatcher, W. W. (2003). Effect of resynchronization with GnRH on day 21 after artificial insemination on pregnancy rate and pregnancy loss in lactating dairy cows. *Theriogenology*, *60*, 1389–1399.
- Christiansen, D. (2015). Pregnancy diagnosis: Rectal palpation. In R. M. Hopper (Ed.), *Bovine reproduction* (3rd ed., pp. 314–319). John Wiley & Sons.
- Colloton, J., DesCôteaux, L., Gayraud, V., & Picard-Hagen, N. (2010). Bovine pregnancy. In J. Colloton, L. DesCôteaux, & G. Gnemmi (Eds.), *Ruminant and camelid reproductive ultrasonography* (1st ed., pp. 81–99). Blackwell Publishing.
- Curran, S., Pierson, R. A., & Ginther, O. J. (1986). Embryonic loss and ultrasonic anatomy of the bovine conceptus on day 10 to 20. *Journal of the American Veterinary Medical Association*, *189*, 1289–1294.
- De Vries, A. (2006). Economic value of pregnancy in dairy cattle. *Journal of Dairy Science*, *89*, 3876–3885.
- Debertolis, L., Mari, G., Merlo, B., Merbach, S., Schoon, H. A., Iacono, E., & Bollwein, H. (2016). Effects of induced endometritis on uterine blood flow in cows as evaluated by transrectal Doppler sonography. *Journal of Veterinary Science*, *17*, 189–197.
- DesCôteaux, L., Colloton, J., & Gnemmi, G. (2009). Ultrasonography of the bovine female genital tract. *Veterinary Clinics of North America Food Animal Practice*, *25*(3), 733–752. <https://doi.org/10.1016/j.cvfa.2009.07.009>
- DesCôteaux, L., Picard-Hagen, N., Durocher, J., Buczinski, S., Colloton, J., Chastant-Maillard, S., & Curran, S. (2010). Bovine fetal development after 55 days, fetal sexing, anomalies, and well-being. In J. Colloton, L. DesCoteaux, & G. Gnemmi (Eds.), *Ruminant and camelid reproductive ultrasonography* (1st ed., pp. 101–124). Blackwell Publishing.
- Díaz, P. U., Belotti, E. M., Notaro, U. S., Salvetti, N. R., Leiva, C. J. M., Durante, L. I., Marelli, B. E., Stangaferro, M. L., & Ortega, H. H. (2019). Hemodynamic changes detected by Doppler ultrasonography in the ovaries of cattle during early development of cystic ovarian disease. *Animal Reproduction Science*, *209*, 106–164.
- Diskin, M. G., Murphy, J. J., & Sreenan, J. M. (2006). Embryo survival in dairy cows managed under pastoral conditions. *Animal Reproduction Science*, *96*, 297–311.
- Faustini, M., Battocchio, M., Vigo, D., Prandi, A., Veronesi, M. C., Comin, A., & Cairoli, F. (2007). Pregnancy diagnosis in dairy cows by whey progesterone analysis: An ROC approach. *Theriogenology*, *67*, 1386–1392.
- Fissore, R. A., Edmondson, A. J., Pashen, R. L., & BonDurant, R. H. (1986). The use of ultrasonography for the study of the bovine reproduction tract. II. Non pregnant, pregnant and pathological conditions of the uterus. *Animal Reproduction Science*, *12*(3), 167–177. [https://doi.org/10.1016/0378-4320\(86\)90037-0](https://doi.org/10.1016/0378-4320(86)90037-0)
- Fricke, P. M. (2002). Scanning the future—Ultrasonography as a reproductive management tool for dairy cattle. *Journal of Dairy Science*, *85*, 1918–1926.
- Fricke, P. M., Ricci, A., Giordano, J. O., & Carvalho, P. D. (2016). Methods for and implementation of pregnancy diagnosis in dairy cows. *Veterinary Clinics of North America: Food Animal Practice*, *32*, 165–180.
- Ginther, O. J. (1995). Luteal glands. In O. J. Ginther (Ed.), *Ultrasonic imaging and animal reproduction: Cattle* (1st ed., pp. 59–71). Equiservices Publishing.
- Ginther, O. J. (2007). Luteal blood flow. In O. J. Ginther (Ed.), *Ultrasonic imaging and animal reproduction: Color-Doppler ultrasonography* (1st ed., pp. 115–148). Equiservices Publishing.

- Giordano, J. O., Guenther, J. N., Lopes, G., Jr., & Fricke, P. M. (2012). Changes in serum pregnancy-associated glycoprotein, pregnancy-specific protein B, and progesterone concentrations before and after induction of pregnancy loss in lactating dairy cows. *Journal of Dairy Science*, *95*, 683–697.
- Gnemmi, G. (2001). La sindrome di degenerazione cistica dell'ovaio nel bovino. *Summa*, *1*, 35–40.
- Gnemmi, G. (2004a). Valutazione ecografica della diagnosi precoce di gravidanza. In G. Gnemmi (Ed.), *Atlante di ultrasonografia ginecologica buiatica* (1st ed., pp. 21–27). Le Point Vétérinaire Italie srl.
- Gnemmi, G. (2004b). Valutazione dei quadri utero-ovarici patologici. In G. Gnemmi (Ed.), *Atlante di ultrasonografia ginecologica buiatica* (1st ed., pp. 67–118). Le Point Vétérinaire Italie srl.
- Gnemmi, G., & Maraboli, C. (2008). Diagnosi ultrasonografica precoce di gravidanza. *Considerazioni pratiche. Summa*, *3*, 1–6.
- Gorjidoz, M., Rahimabadi, P. D., Raofi, A., Masoudifard, M., & Mardjanmehr, S. H. (2021). Ultrasonographic approach and findings in calves with hydranencephaly. *Veterinary Medicine and Science*, *7*, 1718–1727.
- Guimaraes, C. R. B., Oliveira, M. E., Rossi, J. R., Fernandes, C. A. C., Viana, J. H. M., & Palhao, M. P. (2015). Corpus luteum blood flow evaluation on day 21 to improve the management of embryo recipient herds. *Theriogenology*, *84*, 237–241.
- Herzog, K., Brockhan-Lüdemann, M., Kaske, M., Beindorff, N., Paul, V., Niemann, H., & Bollwein, H. (2010). Luteal blood flow is a more appropriate indicator for luteal function during the bovine estrous cycle than luteal size. *Theriogenology*, *73*, 691–697.
- Herzog, K., Voss, C., Kastelic, J. P., Beindorff, N., Paul, V., Niemann, H., & Bollwein, H. (2011). Luteal blood flow increases during the first three weeks of pregnancy in lactating dairy cows. *Theriogenology*, *75*, 549–554.
- Kastelic, J., Curran, S., & Ginther, O. J. (1989). Accuracy of ultrasonography for pregnancy diagnosis on days 10 to 22 in heifers. *Theriogenology*, *31*, 813–820.
- Kito, S., Okuda, K., Miyazawa, K., & Sato, K. (1986). Study on the appearance of the cavity in the corpus luteum of cows by using ultrasonic scanning. *Theriogenology*, *25*(2), 325–333.
- Kraisoon, A., Navanukraw, C., Inthamonee, W., & Bunma, T. (2018). Embryonic development, luteal size and blood flow area, and concentrations of PGF₂ α metabolite in dairy cows fed a diet enriched in polyunsaturated or polyunsaturated fatty acid. *Animal Reproduction Science*, *195*, 291–301.
- Lei, Z. M., Chegini, N., & Rao, C. V. (1991). Quantitative cell composition of human and bovine corpora lutea from various reproductive states. *Reproductive Biology*, *44*, 1148–1156.
- López-Gatius, F., Andreu-Vázquez, C., Mur-Novales, R., Cabrera, V. E., & Hunter, R. H. F. (2017). The dilemma of twin pregnancies in dairy cattle. A review of practical prospects. *Livestock Science*, *197*, 12–16.
- Lüttgenau, J., & Bollwein, H. (2014). Evaluation of bovine luteal blood flow by using color Doppler ultrasonography. *Reproductive Biology*, *14*, 103–109.
- Martins, J. P. N., Wang, D., Mu, N., Rossi, G. F., Martini, A. P., Martins, V. R., & Pursley, J. R. (2018). Level of circulating concentrations of progesterone during ovulatory follicle development affects timing of pregnancy loss in lactating dairy cows. *Journal of Dairy Science*, *101*, 10505–10525.
- McNeill, R. E., Diskin, M. G., Sreenan, J. M., & Morris, D. G. (2006). Associations between milk progesterone concentration on different days and with embryo survival during the early luteal phase in dairy cows. *Theriogenology*, *65*, 1435–1441.
- Meira, E. B., Jr., Henriques, L. C., Sá, L. R., & Gregory, L. (2012). Comparison of ultrasonography and histopathology for the diagnosis of endometritis in Holstein-Friesian cows. *Journal of Dairy Science*, *95*, 6969–6973.
- Miyamoto, A., Shirasuna, K., Wijayagunawardane, M. P. B., Watanabe, S., Hayashi, M., Yamamoto, D., Matsui, M., & Acosta, T. J. (2005). Blood flow: A key regulatory component of corpus luteum function in the cow. *Domestic Animal Endocrinology*, *29*, 329–339.
- Munoz del Real, L. M., Valencia, G. L., Corral, A., & Renteria, T. (2006). Evaluation of ultrasound for pregnancy diagnosis between 20 and 40 days post insemination in dairy cows. *Journal of Animal and Veterinary Advances*, *5*, 949–957.
- Murakami, T., Sato, Y., Sato, A., Mukai, S., Kobayashi, M., Yamada, Y., & Kawakami, E. (2019). Transrectal ultrasonography and blood lactate measurement: A combined diagnostic approach for severe uterine torsion in dairy cattle. *Journal of Veterinary Medical Science*, *81*, 1385–1388.
- Mwaanga, E., & Janowski, T. (2000). Anoestrus in dairy cows: Causes, prevalence and clinical forms. *Reproduction in Domestic Animals*, *35*, 193–200.
- O'Shea, J. D., Rodgers, R. J., & D'Occhio, M. J. (1989). Cellular composition of the cyclic corpus luteum of the cow. *Journal of Reproduction and Fertility*, *85*, 483–487.
- Palhão, M. P., Ribeiro, A. C., Martins, A. B., Guimarães, C. R. B., Alvarez, R. D., Seber, M. F., Fernandes, C. A. C., Neves, J. P., & Viana, J. H. M. (2020). Early resynchronization of non-pregnant beef cows based in corpus luteum blood flow evaluation 21 days after timed-AI. *Theriogenology*, *146*, 26–30.
- Peter, A. T., Vos, P. L., & Ambrose, D. J. (2009). Postpartum anestrus in dairy cattle. *Theriogenology*, *71*, 1333–1342.
- Pierson, R. A., & Ginther, O. J. (1984). Ultrasonography for detection of pregnancy and study of embryonic development in heifers. *Theriogenology*, *22*, 225–233.
- Pugliesi, G., de Melo, G. D., Ataide, G. A., Jr., Pellegrino, C. A. G., Silva, J. B., Rocha, C. C., Motta, I. G., Vasconcelos, J. L. M., & Binelli, M. (2018). Use of Doppler ultrasonography in embryo transfer programs: Feasibility and field results. *Animal Reproduction*, *15*, 239–246.
- Pugliesi, G., Miagawa, B. T., Paiva, Y. N., Franca, M. R., Silva, L. A., & Binelli, M. (2014). Conceptus-induced changes in the gene expression of blood immune cells and the ultrasound-accessed luteal function in beef cattle: How early can we detect pregnancy? *Biology of Reproduction*, *91*, 1–12.
- Pugliesi, G., Rezende, R. G., da Silva, J. C. B., Lopes, E., Nishimura, T. K., Baruselli, P. S., Madureira, E. H., & Binelli, M. (2017). Use of Doppler ultrasonography in timed-AI and ET programs in cattle. *Revista Brasileira de Reprodução Animal*, *41*, 140–150.
- Quintela, L., Barrio, M., Peña, A., Becerra, J., Cainzos, J., Herradón, P., & Díaz, C. (2012). Use of ultrasound in the reproductive Management of Dairy Cattle: Ultrasound in the reproductive management. *Reproduction in Domestic Animals*, *47*, 34–44.
- Ranasinghe, R. M., Nakao, T., Yamada, K., Koike, K., Hayashi, A., & Dematawewa, C. M. (2011). Characteristics of prolonged luteal phase identified by milk progesterone concentrations and its effects on reproductive performance in Holstein cows. *Journal of Dairy Science*, *94*, 116–127.
- Reese, S. T., Pereira, M. H. C., Edwards, J. L., Vasconcelos, J. L. M., & Pohler, K. G. (2018). Pregnancy diagnosis in cattle using pregnancy associated glycoprotein concentration in circulation at day 24 of gestation. *Theriogenology*, *106*, 178–185.
- Ricci, A., Carvalho, P. D., Amundson, M. C., Fourdraine, R. H., Vincenti, L., & Fricke, P. M. (2015). Factors associated with pregnancy-associated glycoprotein (PAG) levels in plasma and milk of Holstein cows during early pregnancy and their effect on the accuracy of pregnancy diagnosis. *Journal of Dairy Science*, *98*, 2502–2514.
- Romano, J. E., Thompson, J. A., Forrest, D. W., Westhusin, M. E., Tomaszewski, M. A., & Kraemer, D. C. (2006). Early pregnancy diagnosis by transrectal ultrasonography in dairy cattle. *Theriogenology*, *66*, 1034–1041.

- Sá Filho, M. F., Marques, M. O., Giroto, R., Santos, F. A., Sala, R. V., Barbuio, J. P., & Baruselli, P. S. (2014). Resynchronization with unknown pregnancy status using progestin-based timed artificial insemination protocol in beef cattle. *Theriogenology*, *81*, 284–290.
- Sauls, J. A., & Stevenson, J. S. (2017). Resynchronization of lactating dairy cows at open pregnancy diagnosis based on the presence or absence of a corpus luteum: A practical approach. *Kansas Agricultural Experiment Station Research Reports*, *3*, 1–9.
- Scully, S., Butler, S. T., Kelly, A. K., Evans, A. C. O., Lonergan, P., & Crowe, M. A. (2014). Early pregnancy diagnosis on days 18 to 21 postinsemination using high-resolution imaging in lactating dairy cows. *Journal of Dairy Science*, *97*, 3542–3557.
- Sharma, A., Singh, M., Abrol, A., & Soni, T. (2019). Doppler sonography of uterine blood flow at mid-oestrus during different degree of clinical endometritis in dairy cows. *Reproduction in Domestic Animals*, *54*, 1274–1278.
- Silva del Rio, N., Colloton, J. D., & Fricke, P. M. (2009). Factors affecting pregnancy loss for single and twin pregnancies in a highproducing dairy herd. *Theriogenology*, *71*, 1462–1471.
- Siqueira, L. G. B., Arashiro, E. K., Ghetti, A. M., Souza, E. D., Feres, L. F., Pfeifer, L. F., Fonseca, J. F., & Viana, J. H. (2019). Vascular and morphological features of the corpus luteum 12 to 20 days after timed artificial insemination in dairy cattle. *Journal of Dairy Science*, *102*, 5612–5622.
- Siqueira, L. G. B., Areas, V. S., Ghetti, A. M., Fonseca, J. F., Palhao, M. P., Fernandes, C. A. C., & Viana, J. H. M. (2013). Color Doppler flow imaging for the early detection of nonpregnant cattle at 20 days after timed artificial insemination. *Journal of Dairy Science*, *96*, 6461–6472.
- Stroud, B. K., & Durocher, J. (2010). Bovine embryo transfer, in vitro fertilization, special procedures, and cloning. In J. Colloton, L. DesCoteaux, & G. Gnemmi (Eds.), *Ruminant and camelid reproductive ultrasonography* (1st ed., pp. 125–142). Blackwell Publishing.
- Szenci, O. (2021). Recent possibilities for the diagnosis of early pregnancy and embryonic mortality in dairy cows. *Animals*, *11*(6), 1666–1683.
- Szenci, O., Varga, J., & Bajcsy, A. C. (1999). Role of early pregnancy diagnosis by means of ultrasonography in improving reproductive efficiency in a dairy herd: A retrospective study. *The Bovine Practitioner*, *33*(1), 67–69.
- Utt, M. D., Johnson, G. L., & Beal, W. E. (2009). The evaluation of corpus luteum blood flow using color-flow Doppler ultrasound for early pregnancy diagnosis in bovine embryo recipients. *Theriogenology*, *71*, 707–715.
- Viana, J. H. M., Arashiro, E. K. N., Siqueira, L. G. B., Ghetti, A. M., Areas, V. S., Guimaraes, C. R. B., Palhao, M. P., Camargo, L. S. A., & Fernandes, C. A. C. (2013). Doppler ultrasonography as a tool for ovarian management. *Animal Reproduction*, *10*, 215–222.
- Wijma, R., Stangaferro, M. L., Masello, M., Granados, G. E., & Giordano, J. O. (2017). Resynchronization of ovulation protocols for dairy cows including or not including gonadotropin-releasing hormone to induce a new follicular wave: Effects on re-insemination pattern, ovarian responses, and pregnancy outcomes. *Journal of Dairy Science*, *100*, 7613–7625.

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